

AMENDMENTS TO THE SPECIFICATION

Please replace paragraph 1 with the following amended paragraph 1:

-- The present invention relates to the inhibitor of angiogenesis comprising ~~tetraacetylphytosphingosine~~ tetraacetylphytosphingosine derivatives and the kit for treating cancer comprising the inhibitor.

Please replace paragraph 14 with the following amended paragraph 14:

--For the kit for treating cancer, the spingolipid derivatives are one or more spingolipid derivatives selected from the group consisting of phytosphingosine, ~~enacethylphytosphingosine~~ N-acetylphytosphingosine, C6 phytoceramide, C8 phytosphingosine, dimethylsphingosine, dimethylphytosphingosine and sphingosine.--

Please replace paragraph 20 with the following amended paragraph 20:

-- FIG. 5 A-C is are photographs ~~photographes~~ representing a result of angiogenesis ~~angiognesis~~-test of a solution containing tetraacetylphytosphingosine according to the present invention. --

Please replace paragraph 21 with the following amended paragraph 21:

--FIG. 6 A-D is are photographs ~~photographes~~ representing that a solution containing tetraacetylphytosphingosine according to the present invention inhibited migration of endothelial cells. --

Please replace paragraph 39 with the following amended paragraph 39:

-- Female New Zealand White rabbits (body weight 2.0 kg) were used as experimental animals in this assay. PBS solution containing 0.1% BSA (BSA-PBS solution) was prepared as control solution for negative control group (comparative example 1). ~~sphingosylphosphorylcholine~~

Sphingosylphosphorylcholine (comparative example 2), phytosphingosine (comparative example 3), ~~N-acetylphytosphingosine~~ acetylphytosphingosine (comparative example 4) as positive control group, and tetraacetyl phytosphingosine (example 1) were dissolved in ethanol or methanol respectively. Each portion of the solutions was added to silicone glass tube. Then, it was charged with N₂ gas, and added 0.1% BSA-PBS solution. After coupling them with water sonicator and vortex, these solutions were spotted on and injected intradermally to wound region of experimental animals. At the same time, to study an effect of the angiogenesis as concentrations of ~~tetraacethylsphingosine~~ tetraacetylsphingosine, the animal was treated with 0.1 μM, 1 μM, 2 μM and 5 μM of ~~tetraacethylsphingosine~~ tetraacetylsphingosine, and ~~compared~~ compared with the results. Experimental animals were put into special stainless cages which was designed suitably to assay experimental animals, anesthetized by injecting ketanine (3-4 mg/kg) intramuscularly. The hair and corneous tissue of innerside of both ears were removed with shaving and washing, and then were disinfected with 70% ethanol. Four wound regions per ear were formed by using 6 mm punch for sldn histological examination (Stiefel, Germany) under sterile condition if possible, and each wound region was spotted or injected intradennally 30-50 μl of the control solution or each treatment material. Wound regions were sealed with cathereep (Nichiban Co., Tokyo Japan) which was cut in size which are greater than that of wound region to prevent contamination of the wound regions and formation of crust. Then, the wound regions were protected with 2.times.2 gauze, and ears of rabbit were bandaged with elastopore (Nichiban Co., Tokyo Japan). Subsequently, the rabbits were bred in a cage per a rabbit. After 48 hrs, same procedure was repeated. On 4 days and 8 days after forming wound regions, sacrificing the rabbits and treating tissue for histological study. For histological study, wound tissues were fixed with 10% formalin, cut it in half longitudinally and made paraffin block. Then, approximately 5 μm of segment was made, attached it to slide and stained with hematoxylin and eosin to observe a change of epidermis and dermis, and stained with Massons Trichrome to observe a degree of collagen formation of granulation tissue.--

Please replace paragraph 41 with the following amended paragraph 41:

-- FIG. 1 is a graph representing the number of blood vessels determined on 4 days after treatment of 0.1 μM, 1 μM, 2 μM and 5 μM of a solution containing tetraacetylphytosphingosine according to

the present invention respectively in comparison with negative control. In FIG. 1, C-4 represents a result of a negative control group which elapsed 4 days after treatment. T-0.1-4 represents a result of a experimental group which elapsed 4 days after treatment of 0.1 μ M of ~~tetraacetylphytosphingosine~~ tetraacetylphytosphingosine (TAPS). T-1-4, T-24 and T-54 represent results of experimental groups which elapsed 4 days after treatment of 1 μ M, 2 μ M and 5 μ M of TAPS respectively. As shown in FIG. 1, ~~tetraacetylphytosphingosine~~ tetraacetylphytosphingosine inhibited angiogenesis greatly. --

Please replace paragraph 43 with the following amended paragraph 43:

-- Meanwhile, same experiments were carried out with 5 μ M of the negative control group (comparative example 1), sphingosylphosphorylcholine (SPC)(comparative example 2), phytosphingosine (PS)(comparative example 3), ~~N-acetylphytosphingosine~~ acetylphytosphingosine (NAPS)(comparative example 4) and tetraacetylphytosphingosine (TAPS) as example (example 1) respectively, then the number of blood vessels and the area of granulation tissue were determined. The results were as follows:--